$Dl_{i} = l_{i} = l_{i} = l_{i}$ 

# Hydraulic architecture of seedlings and adults of Rhizophora mangle L. In fringe and scrub mangrove

# Arquitectura hidráulica de plántulas y adultos de Rhizophora mangle L. en manglar chaparro y de franja

DIANA J. CISNEROS-DE LA CRUZ<sup>1</sup>, LAURA YÁÑEZ-ESPINOSA<sup>2</sup>, CASANDRA REYES-GARCÍA<sup>1</sup>, ROBERTH US-SANTAMARIA<sup>1</sup>, JOSÉ LUIS ANDRADE<sup>1\*</sup>

#### Abstract

**Background:** Mangrove plant species have distinctive anatomical and physiological responses to cope with a wide range of salinities and inundations. These strategies pertain a safe and efficient water use and transport, essential for survival.

**Questions:** How are the anatomical and physiological attributes of the hydraulic architecture of seedlings and adults of *Rhizophora mangle*? what are the changes in hydraulic architecture of seedlings and adults of *R. mangle* in contrasting microenvironments?

Studied species: Rhizophora mangle L. (Rhizophoraceae).

Study site and dates: Scrub and fringe mangroves in Ria Celestún Biosphere Reserve, during the rainy season of 2013 (July to October).

**Methods:** Hydraulic conductivity and leaf water potential, as well as xylem vessel density, length, transversal and radial diameter, and area were measured for seedlings and adults from both sites. The prevailing environmental conditions (soil water potential, salinity, photon flux density, air temperature and relative humidity) were also characterized.

**Results:** A safer hydraulic conduction system, with narrow and more grouped vessels, was observed in seedlings than in adults of *R. mangle* in both sites. Adult individuals from the scrub mangrove, in the hyper saline microenvironment, had a safer hydraulic conduction system than adults in the fringe mangrove.

**Conclusions:** The seedling stage of *R. mangle* showed a safer hydraulic system than adults in both types of mangroves. However, over time this hydraulic conduction system could become more efficient or remain safe depending on the microenvironment in which individuals are growing. **Keywords:** Hydraulic conductivity, mangrove type, salinity, water potentials, Yucatan.

#### Resumen

**Antecedentes:** Los manglares presentan respuestas anatómicas y fisiológicas distintivas para enfrentar una amplia variación de condiciones ambientales. Éstas están relacionadas con la seguridad y eficiencia en el uso y transporte del agua para su supervivencia.

**Preguntas:** ¿Cómo son los atributos anatómicos y fisiológicos de la arquitectura hidráulica de las plántulas y los adultos de *R. mangle*? ¿Cuáles son los cambios en la arquitectura hidráulica de plántulas y adultos de *R. mangle*, en microambientes contrastantes?

Especies de estudio: Rhizophora mangle L. (Rhizophoraceae).

Sitio y años de estudio: Manglares chaparros y de franja en la Reserva de la Biósfera Ria Celestún, durante la estación lluviosa de 2013.

**Métodos:** Se realizaron mediciones de la conductividad hidráulica y el potencial hídrico foliar, así como aspectos anatómicos de los vasos del xilema, en plántulas y los adultos de ambos sitios. También se caracterizaron las condiciones ambientales (potencial hídrico del suelo, salinidad, densidad de flujo de fotones, temperatura del aire y humedad relativa del aire).

Resultados: El sistema de conducción hidráulica de las plántulas de ambos sitios fue más seguro, con vasos estrechos y más agrupados, que en adultos. Los adultos del manglar chaparro tuvieron un sistema de conducción más seguro que en los adultos del manglar de franja.

Conclusiones: Las plántulas de *R. mangle* mostraron un sistema de conducción hidráulico más seguro que los adultos en ambos tipos de manglar. Sin embargo, con el tiempo, el sistema de conducción puede revertir en un sistema más eficiente o permanecer seguro, dependiendo del microambiente en el que se desarrollen los individuos.

Palabras clave: Conductividad hidráulica, potenciales hídricos, salinidad; tipo de manglar, Yucatán.





<sup>&</sup>lt;sup>1</sup> Unidad de Recursos Naturales, Centro de Investigación Científica de Yucatán, Mérida, México.

<sup>&</sup>lt;sup>2</sup>Instituto de Investigación de Zonas Desérticas, Universidad Autónoma de San Luis Potosí, San Luis Potosí, México.

<sup>\*</sup>Author for correspondence: andrade@cicy.mx

angrove forests grow in the intertidal zone on tropical and subtropical coasts (Lugo & Snedaker 1974, Feller & Sitnik 1996). Consequently, mangrove species can tolerate a wide range of salinities and flooding that, together with other factors, such as topography, soil physiochemistry and mineral nutrients, result in diverse types of forests with different canopy height and species composition (Lugo & Snedaker 1974, López-Portillo & Ezcurra 2002). For instance, in the Yucatan Peninsula, which host 54.5 % of the total area occupied by mangroves in Mexico (Rodríguez-Zúñiga *et al.* 2013), there are different mangrove types, such as fringe, basin, scrub, and peten (Zaldívar-Jiménez *et al.* 2010). In all types of mangrove forests, *Rhizophora mangle* L. can occur with a contrasting structure: tall trees in the less saline fringe mangrove and shorter trees in the hypersaline scrub mangrove (Zaldívar-Jiménez *et al.* 2004). Scrub mangrove plants are supposedly short because of several complex abiotic factors such as salinity, flooding, redox potentials, and mineral nutrient availability (Lugo & Snedaker 1974, Feller *et al.* 2003, Lovelock *et al.* 2006), but data of salinity and phosphorous in sediments presented in several tall and scrub mangrove forests are contradictory (Cisneros-de la Cruz *et al.* 2018).

The different types of mangroves show specific adaptive strategies, according to the environmental conditions in which they grow (Feller 1996, Robert *et al.* 2009, Yáñez-Espinosa & Flores 2011). For example, some studies have shown that scrub mangrove have less photosynthetic efficiency, gas exchange and hydraulic conductivity than fringe mangroves (Lin & Sternberg 1992, Lovelock *et al.* 2006, Naidoo 2010). This occurs because, in the hypersaline environment, plants experience long periods of physiological drought (Yáñez-Espinosa *et al.* 2001, López-Portillo *et al.* 2005, Schmitz *et al.* 2006a). In any case, one of the most critical tradeoffs is to reconcile efficient water transport with security against cavitation, which means the obstruction of the conducts because of the formation of air bubbles (Schmitz *et al.* 2006b, Xiao *et al.* 2009). Then, study of the hydraulic architecture of this mangrove species would improve the understanding on the mechanisms of water movement, under extreme saline environments, and its responses to environmental changes (Tyree & Ewers 1991, Cruiziat *et al.* 2002, McCulloh & Sperry 2005).

Adaptations in hydraulic architecture in mangrove trees can be key in regulating their distribution, survival, and growth (Tyree & Ewers 1991, Valladares *et al.* 2004); and are especially relevant at the seedling stage when individuals are more vulnerable to environmental variability (Cornelissen *et al.* 1996, Ishida *et al.* 2005, Marks 2007, Krauss *et al.* 2008). However, only one study has focused on the physiological and anatomical strategies during different life stages (Farnsworth & Ellison 1996).

Since mangroves are constantly exposed to anthropic and natural impacts that rapidly diminish their natural distribution (Alongi 2002), the characterization of hydraulic architecture strategies from seedlings to adults under contrasting environments could strengthen management actions toward their restoration and conservation. It is expected that scrub mangroves growing under a more saline environment will exhibit more secure hydraulic strategies than fringe mangroves at both life stages, seedling, and adult. The objective of this study was to compare the anatomical and physiological attributes of stem hydraulic architecture of seedlings and adults of *Rhizophora mangle* L. in the contrasting environments of a scrub and fringe mangrove forests in Celestún, Yucatan, Mexico.

### Materials and methods

Study area and microenvironment. The study was performed in the Ria Celestún Biosphere Reserve located at the northwest of the Yucatan Peninsula (Figure 1). The climate is warm semi-arid with a mean annual temperature of 28.5 °C, annual mean precipitation of 760 mm, and characterized by three seasons: dry (March-May), rainy (June-October) and early dry (locally named "nortes"; showing dispersed rain events, November-February) (Zaldívar-Jiménez *et al.* 2010). The tidal regime characteristic is a mixed semidiurnal tide of 0.6 m (Herrera-Silveira 1994). Two sampling points were chosen and correspond to two types of mangroves with contrasting morphology and environmental conditions; their location was based on the study area of Zaldívar-Jiménez *et al.* (2004) and on sites included in the Mexico Mangrove Monitoring System (Herrera-Silveira *et al.* 2014). The fringe mangrove (Figure 1A: 20° 51' 22.7" N; 90° 22' 37.7" W) was located at the edge of the lagoon, where trees were  $16.3 \pm 4.8$  m tall, and had a diameter at breast height (dbh) of  $22.9 \pm 7.2$  cm, and a density of 1,583 trees ha<sup>-1</sup>. The scrub mangrove (Figure 1B:  $20^{\circ}$  51' 04.9" N;  $90^{\circ}$  22' 22.9" W) was located 0.72 km inland from the lagoon, where trees were  $2.8 \pm 1$  m tall, had a dbh of  $3.4 \pm 1.5$  cm, and a density of 15,600 trees ha<sup>-1</sup>.

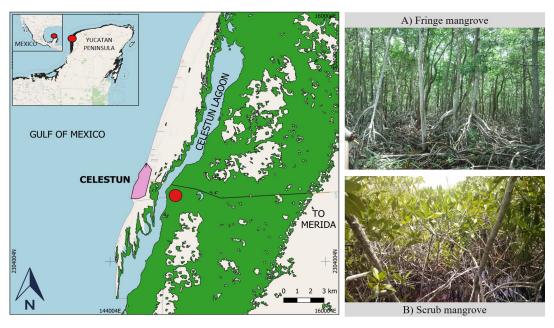


Figure 1. Location of the study site and mangrove forests in the Ria Celestún Biosphere Reserve, Yucatan, Mexico.

Rhizophora mangle L. is a viviparous angiosperm, with high ecological importance, which shows great plastic phenotype and can grow in a range of salinity of from 0 to 90 ‰ (Tomlinson 2016). For the two-mangrove type selected, two plant stages of *R. mangle* were considered: seedlings, individuals with visible propagule of < 50 cm height and diameter < 1 cm, with 10 - 20 leaf scars to ensure similar ages (Duke & Pinzon 1992); and adults, individuals from 2 to 4 m height and from 2.5 to 3.5 cm dbh. Sampling was achieved within a three-month period during the rainy season.

Microenvironment was simultaneously characterized in the understory at both sites and on one reference point above the canopy during a week in July, August, and October 2013. Air temperature and relative humidity were recorded with a Temp Smart Sensor (S-TMB-M003, Onset Computer Corporation, Bourne, MA), and photosynthetic photon flux (PPF) with a quantum sensor (Photosynthetic Light Smart Sensor, S-LIA-M003, Onset, Bourne, MA). Variables were recorded simultaneously every 10 s and 10-min averages were stored with a data acquisition system (HOBO U30-NRC Weather Station Starter Kit, Onset, Bourne, MA), such interval allows to save representative information of the microenvironmental changes without saturating the datalogger memory. Vapor pressure deficit (VPD) was estimated after Jones (2013). During each microenvironment measurements, interstitial water was sampled with an acrylic tube connected with a syringe to measure interstitial salinity ( $\sim$  30 cm depth) using a portable conductivity meter (Model 30, YSI, Yellow Springs, OH) at three random points per site. At each point, one soil sample were collected at equal depths with a PVC tube and a plunger, stored on ice within plastic bags and carried to the laboratory where soil water potential ( $\Psi_{soil}$ ) was measured with a dew point potentiometer (WP4, Decagon Devices Inc., Pullman, WA).

Physiological measurements. For all plant measurements, five individuals of R. mangle, per life stage and site, were considered. Three fully developed leaves were collected from every individual to measure water potential at pre-dawn (maximum water potential;  $\Psi_{predawn}$ ) and noon (minimum water potential;  $\Psi_{midday}$ ), with a dewpoint potentiometer (WP4, Decagon Devices Inc., Pullman, WA). To measure the hydraulic conductivity ( $k_h$ ; kg m<sup>-1</sup> MPa<sup>-1</sup>), individuals were removed, covered with wet towels and plastic. Segments 20 cm long were measured with a hydraulic conductance and flow meter in the transient mode (HCFM, Dynamax, Houston, TX), according to Sperry et al. (1988). The specific hydraulic conductivity ( $k_s$ ; kg m<sup>-1</sup> s<sup>-1</sup> MPa<sup>-1</sup>) was obtained as the ratio between  $k_h$  and xylem area (Melcher et al. 2001).

Anatomical characterization. After measuring the hydraulic conductivity, samples for each stage and site were fixed in FAA (formaldehyde, ethanol 96°, glacial acetic acid, water 10: 35: 5: 50) (Ruzin 1999); and, after two days, washed with tap water and stored in GAA (glycerin, 96° ethanol, water 1: 1: 1) until sectioning. Subsequently, smaller segments were extracted from the center of the samples and using a sliding microtome (GSL1, Swiss Federal Research Institute WSL, Birmensdorf, Switzerland), 50 µm thick transverse sections were obtained. Tissue sections were stained with safranin-fast green (Ruzin 1999) and mounted in permanent slides with synthetic resin. Sections were analyzed with a light microscope (DM2000, Leica, Wetzlar, Germany) and digital images of tree fields for each slide were taken with an integrated camera Olympus U-CMAD3, equipped with the program Infinity Analyze (release 5.0.2; Lumenera Corp., Canada). The measurements were carried out with the ImageJ (v1.49c; National Institutes of Health, USA) in three visual fields of each image captured. The tangential and radial diameter of the vessel lumen, the thickness of the vessel wall and vessel density were measured. The area of the vessel lumen (VLA) was calculated using the equation for an oval area as  $VLA = \pi x^{td} \frac{v}{2} x^{rd} \frac{v}{2}$ , where td = vessel tangential diameter and rd = vessel radial diameter. The percentage of solitary vessels and vessels arranged in radial multiples and clusters, were also obtained. The length of the vessel elements was measured in wood macerated using Jeffrey's solution (Ruzin 1999).

Data analysis. Nested analyses of variance were performed to compare the means of the anatomical and physiological variables between life stages and sites; stage was considered nested within mangrove type. Duncan post-hoc tests were done to identify significant differences among means (P < 0.05). Shapiro-Wilk's tests were used to assess whether data distribution was normal, and Levene's tests were used to test for homoscedasticity. Data without normal distribution or homoscedasticity were transformed with natural logarithm or reciprocal transformations. When normal distribution for the data was not achieved (grouping vessels) a Kruskal-Wallis test was performed. All analyzes were performed in STATISTICA 7 (StatSoft, Tulsa, USA). To distinguish the variables that contribute the most to the variance of groups, a canonical discriminant analysis (CDA) was performed using the physiological and anatomical variables that did not present multicollinearity (vessel density, lumen area and length of the vessel, % of solitary vessels, specific hydraulic conductivity, and maximum and minimum leaf water potential). Also, a canonical correlation analysis (CCA) was performed with the most relevant discriminant variables; the physiological and anatomical variables as dependent variables (vessel density, specific hydraulic conductivity, vessel area, and maximum and minimum leaf water potential), and the microenvironmental variables (salinity and maximum VPD) as independent variables. The multivariate analyzes were performed with XLSTAT v7.5.2 (Addinsoft, France).

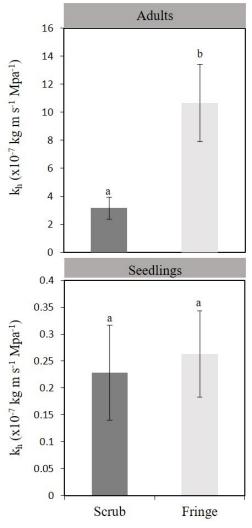
## Results

*Microenvironment*. Fringe and scrub sites showed contrasting microenvironments during the study period. The fringe mangrove site had lower average salinity (25.84  $\pm$  2.75 %) and higher soil water potential ( $\Psi_{soil}$ ; -0.30  $\pm$  0.2 MPa) than the scrub mangrove site (51.42  $\pm$  2.28 % and -1.91  $\pm$  0.94 MPa, respectively) (P < 0.05). Maximum vapor pressure deficit (VPD) in the understory was lower in the fringe mangrove site (1.38  $\pm$  0.12 kPa) than in the scrub mangrove site (1.83  $\pm$  0.12 kPa; P < 0.05). Also, the understory of the fringe site received less daily photosynthetic photon flux density (PPFD; 2.28  $\pm$  0.2 mol m<sup>-2</sup> d<sup>-1</sup>) than that of the scrub site (5.85  $\pm$  0.54 mol m<sup>-2</sup> d<sup>-1</sup>). The reference point above the canopy received up to ten times more PPFD than the understory of both sites (45.2  $\pm$  2.46 mol m<sup>-2</sup> d<sup>-1</sup>). However, VPD values in the scrub site were like those registered above the canopy (Table 1).

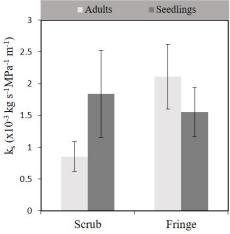
Physiological responses. Hydraulic conductivity  $(k_h)$  was higher for adults in the fringe forest  $(1.06 \pm 0.3 \text{ kg m s}^{-1} \text{ MPa}^{-1})$  than that for adults in the scrub mangrove  $(0.32 \pm 0.07 \text{ kg m s}^{-1} \text{ MPa}^{-1})$  ( $F_{1,2} = 6.93, 14.13; P < 0.05; Figure 2$ ). Differences in  $k_h$  were also found between life stages in the fringe forest (P < 0.05). Specific hydraulic conductivity  $(k_s)$  of plants was not different between life stages within each forest nor between forest types  $(F_{1,2} = 1.7, 0.87; P > 0.05)$ , but fringe adults, and fringe and scrub seedlings, had a tendency of higher  $k_s$  values than shrub adults (Figure 3). Although the  $\Psi_{\text{predawn}}$  was not different for plants at any site or stage, the  $\Psi_{\text{midday}}$  of scrub adults was the lowest (Duncan test, P < 0.05; Figure 4).

**Table 1.** Microenvironment of scrub and fringe mangrove understory and above the canopy in Celestún, Yucatan, Mexico. Data are means ± standard errors.

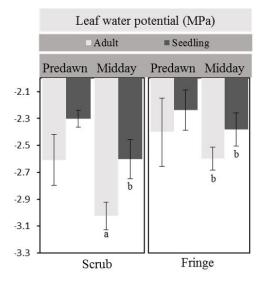
Mangrove type	Temperature (°C)	Relative Humidity (%)	Photosynthetic photon flux density (PPFD) (mol m <sup>-2</sup> d <sup>-1</sup> )	Mean vapor pressure deficit (VPD) (kPa)	Soil water potential (Ψ <sub>soil</sub> ) (MPa)	Interstitial salinity (‰)
Scrub	$27.7 \pm 3.2$	$76.8 \pm 9.5$	$5.8 \pm 0.54$	$0.94 \pm 0.57$	$-1.91 \pm 0.94$	$51.42 \pm 2.28$
Fringe	$27.0 \pm 2.5$	$79.5 \pm 7.5$	$2.28 \pm 0.2$	$0.78 \pm 0.41$	$-0.30 \pm 0.2$	$25.84 \pm 2.75$
Above the canopy	$27.5 \pm 3.4$	$76.9 \pm 10.5$	$45.2\pm2.46$	$0.94 \pm 0.61$	-	-



**Figure 2.** Hydraulic conductivity  $(k_h)$  of seedlings and adults of *Rhizophora mangle* in scrub and fringe mangrove. Different letters indicate significate differences between sites and stage (P > 0.05). Data are means  $\pm$  standard errors (bars indicate standard errors).



**Figure 3.** Specific hydraulic conductivity  $(k_s)$  of seedlings and adults of *Rhizophora mangle* in scrub and fringe mangrove (P < 0.05). Data are means  $\pm$  standard errors (bars indicate standard errors).

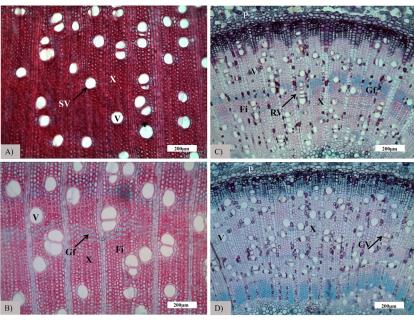


**Figure 4.** Predawn and midday values of foliar water potential of seedlings and adults of *Rhizophora mangle* from fringe and scrub mangrove. Different letters indicate significant differences in midday leaf water potentials between stages and sites (P < 0.05). Data are means  $\pm$  standard errors (bars indicate standard errors).

Anatomical traits. Adults and seedlings of R. mangle had different anatomical stem structures. At seedling stage, stems showed secondary growth without significant differences in vessels characteristics between scrub and fringe mangrove forest (Figure 5; Table 2). Seedlings had an average vessel density of  $103 \pm 35 \text{ v/mm}^2$ , with a tangential and radial diameter of  $37.75 \pm 6.53 \mu m$  and  $41.83 \pm 7.55 \mu m$ , respectively; vessels were mostly solitary (65.43 %) and vessel clusters less frequent (12.35 %). Compared to adults, seedlings had differences in anatomical characteristics related to the allometric relationship of their respective growth stages (Figure 5), smaller vessels, but in a greater density than in adults (Table 2). Adults in fringe mangrove had lower vessels density ( $94 \pm 6 \text{ v/mm}^2$ ) than adults in scrub forest ( $111 \pm 7 \text{ v/mm}^2$ ) (P > 0.01). Also, the vessel grouping pattern observed was different between forest types, with a higher percentage of solitary vessels in the fringe, and higher radial multiple vessels in the scrub (Table 2). Furthermore, vessel lumen area and radial diameter were higher for adults in the fringe than in the scrub mangrove ( $F_1 = 16.5, 6.4, P < 0.05$ ).

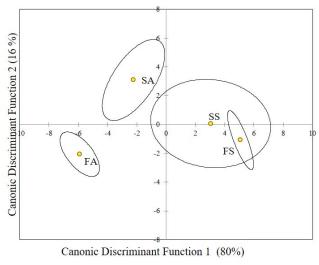
**Table 2.** Anatomical vessel variables for seedlings and adults of *Rhizophora mangle* from the scrub and fringe mangrove. Data are means  $\pm$  standard errors. Different letters indicate significant differences within rows (Duncan test, P < 0.05).

VariableS	Sc	rub	Fringe	
	Adult	Seedling	Adult	Seedling
Vessel density (v/mm²)	$32\pm1$ a	$934\pm6$ b	$19\pm1$ a	111 ± 7 <sup>b</sup>
Tangential diameter (µm)	$72.1\pm2.3$ a	$37.8 \pm 1.0$ b	$84.0 \pm 2.2$ a	$37.7 \pm 1.3$ b
Radial diameter (µm)	$70.4\pm3.8$ b	$41.1 \pm 1.4$ °	$89.7 \pm 3.8$ a	$42.5 \pm 1.4$ °
Wall thickness (µm)	$6.0 \pm 0.2$ a	$3.8\pm0.1$ b	$5.3 \pm 0.2$ a	$3.7 \pm 0.1 \ ^{ab}$
Lumen area (µm²)	$3807 \pm 178$ $^{\rm b}$	$1122 \pm 60^{\circ}$	$5609 \pm 284~^a$	$1136 \pm 68$ °
Length (µm)	$905.5 \pm 26.0$ a	$611.0 \pm 16.1$ b	926.1 ± 21.2 a	$622.6 \pm 17.9$ b
% Solitary vessels	$54.5\pm3.7$ a	$70.7 \pm 4.8~^{\rm ab}$	$72.2 \pm 5.1$ b	$58.8 \pm 4.7 ~^{\rm ab}$
% Cluster's vessels	$14.7\pm3.1~^{\rm a}$	$10.7\pm3.6$ a	$11.9 \pm 3.8$ a	$14.0\pm3.5$ a
% Radial multiples vessels	$31.6 \pm 4.2$ a	$18.5\pm4.5~^{ab}$	$15.9 \pm 4.3$ b	$30.0 \pm 4.5~^{ab}$



**Figure 5.** Transversal sections of stems of seedlings and adults of *Rhizophora mangle* in fringe and scrub mangrove: A) adult scrub; B) adult fringe; C) seedling scrub; and D) seedling fringe (V, vessels; X, xylem; P, phloem; Fi, fibers; Gf, Gelatinous fibers; SV, solitary vessels; RV, vessel radial multiples; CV, vessel clusters).

Multivariate analysis. According to the CDA, the first discriminant function explained 80 % of the total variance and the second 16 % of the remaining variation. The first function explained the differences in adult and seedling stages defined by the vessel density, vessel lumen area, and  $k_s$ ; while the second explained the differences between fringe and scrub mangrove forest through the  $\Psi_{predawn}$  and  $\Psi_{midday}$  (Figure 6) (Wilk's  $\lambda$  = 0.003, n = 120, P < 0.005). The correlation of environmental, physiological, and anatomical variables in the CCA was explained with 62 % in the first canonic correlation (0.94, P < 0.001). The canonical correlation analysis also showed a negative relationship between hydraulic conductivity and vessel lumen area with salinity (Table 3); also, it showed that  $\Psi_{midday}$  and vessel density were negatively related to VPD.



**Figure 6.** Canonical Discriminant Analysis for anatomical and physiological variables of seedlings and adults of *Rhizophora mangle* in fringe and scrub mangrove forests. Wilk's  $\lambda = 0.003$ , P > 0.005. FA: fringe adults; SA: scrub adults; FS: fringe seedlings; SS: Scrub seedlings.

**Table 3.** Anatomical and physiological variables used in the canonical discriminant analysis and their partial contribution expressed by the standardized coefficients of the canonical functions. Letters represent the variables with more contribution in centroids separation.

Variables	Correlation of original and ca- nonical variables	Standardized canonical coefficients	
Anatomical			
Vessel density	0.63	0.11	
Lumen vessel area	-0.82	-0.88 a	
Physiological			
Specific hydraulic conductivity (k <sub>s</sub> )	-0.82	-0.19	
Predawn leaf water potential ( $\Psi_{predawn}$ )	0.03	-0.34	
Midday leaf water potential $(\Psi_{midday})$	-0.10	-0.49	
Environmental			
Maximum vapor pressure deficit ( $VPD_{max}$ )	-0.33	-0.97	
Salinity	0.59	1.13 a	

### Discussion

Seedlings and adults of *Rhizophora mangle* showed physiological and anatomical strategies in the hydraulic conduction system that allowed them to respond to the contrasting environments in which they developed. While the fringe mangrove is in continuous contact with the water body, under salinity values close to those of the lagoon; the scrub mangrove, in the inland zone, has higher and variable salinity values, which are regulated by the sporadic high tide and rain events (Lugo *et al.* 2007, Herrera-Silveira 1994). This explained the differences in salinity and  $\Psi_{\text{soil}}$  of both sites (Table 1). The higher daily photosynthetic photon flux density (PPFD) in the understory of the scrub mangrove, compared to the fringe mangrove, also caused higher temperature and vapor pressure deficit (VPD) values, which were closer to the reference value above the canopy (Table 1).

In these contrasting environments, salinity explained most of the variance in hydraulic architecture of both, seed-ling, and adult stages, within the canonical correlation analysis (CCA). This agrees with previous studies showing that salinity is one of the most important factors influencing the health, distribution, growth, and productivity of mangroves (Ball 2002, Krauss *et al.* 2008). In scrub mangroves, the high salinity, lower soil water potential, and high vapor pressure deficit exert negative pressures on the hydraulic systems that guarantee the water flow from roots to leaves (Jones 2013, Reef & Lovelock 2015), which is reflected in the inverse relation of VPD and the  $\Psi_{\text{midday}}$  in the CCA (Table 3). However, although this increase in pressure within the vessel could form air bubbles that lead to the cavitation vessel in the water column and the consequent reduction of hydraulic conductivity (Sobrado 2007), the narrower vessels and thicker walls, observed in scrub mangroves (Figure 5), would offer greater mechanical strength to resist strong negative pressures (Baas *et al.* 1983, Guet *et al.* 2015). In fact, salt exclusion mangroves, as *R. mangle* have the highest cavitation resistance reported for angiosperm trees (Jiang *et al.* 2017).

The maintenance of the water column continuity is crucial to ensure water supply to leaves and allowing carbon gain for plant growth and survival (Tyree 2003). It has been observed that larger stems are related to wider vessels that ensure hydraulic efficiency through the water column (Rosell et al. 2017). This relation was also observed in this study: wider vessels in fringe adults, with larger stems and water columns, than in shorter stems of individuals of the scrub mangrove. Despite vessel length did not have significant differences between mangrove types (Table 2), a tendency was observed with shorter vessels in scrub mangrove that allow a safer water transport system, and larger vessels in fringe mangrove that resulted in a more efficient water transport (Cruiziat et al. 2002, Gil-Pelegrín et al. 2005). Moreover, the higher vessel density and grouping observed in scrub adults and seedlings of both mangrove types, showed in Figure 5, would allow greater redundancy and, therefore, greater safety against embolisms (Cruiziat et al. 2002, Ewers et al. 2007). These characteristics provide safer hydraulic conductivity but imply a tradeoff with an efficient water flow capacity of each vessel according to the Hagen-Poiseuille law (Tyree & Zimmermann 2002). This inverse relation of hydraulic conductivity with vessel lumen area and salinity was sustained by the CCA (Table 3). Conversely, the low density, wide vessels, and thin vessel walls of fringe mangroves, would allow a more efficient water transport, but high vulnerability to vessel cavitation. These results agree with Lin & Sternberg (1992), who recorded lower CO, assimilation and higher water use efficiency on scrub mangroves than fringe mangroves. Yet, seedlings of both types of mangroves had similar hydraulic architecture, with high density, smaller and more clustered vessels that confer a safer, but a less efficient system, with hydraulic conductivity  $(k_h)$  values well below those of adults (Figures 2, 5).

The lower hydraulic conductivity in seedlings compared to adults highlight the relevance of a safe conducting system at this life stage, even if seedling establishment is generally favored by a greater growth and biomass production (Donovan & Ehleringer 1991). However, a safer conducting system requires higher carbon investment, which implies an additional cost, diminishing growth rates (Ball *et al.* 1997, Sobrado & Ewe 2006, Jiang *et al.* 2017). Low hydraulic conductivity is associated to low photosynthetic rates, concordant with the low stomatal conductance observed in scrub mangroves (Ball & Farquhar 1984, Shiau *et al.* 2017). Moreover, this safer hydraulic system could provide seedlings a greater chance of survival in a variable saline environment until the reproductive stage (Donovan & Ehleringer 1991). Although it has been suggested that seedlings show high plasticity (Farnsworth & Ellison 1996,

Biber 2006), in this study no differences were observed in the hydraulic architecture at this stage despite significant microenvironmental differences in the sites they grew. This could be explained by the viviparity of *R. mangle* (Tomlinson 2016): the propagule is an important carbohydrate reservoir, which the seedlings can use differentially during the first stage of growth according to the specific conditions in which they develop (Smith & Snedaker 2000). Consequently, storage reserves are temporarily available for propagules to buffer from either flooding, salinity and/or variability in resource availability that may otherwise affect photosynthesis and growth (Smith & Snedaker 2000, Dissanayake *et al.* 2014, Lechthaler *et al.* 2016).

One of the changes in water transport strategies related to the growth stage of *R. mangle* is the differences in leaf minimum water potential, as the canonical discriminant analysis (CDA) showed (Figures 4, 6). The leaf water potential of adults at both sites was contrasting, with lower values in scrub compared to fringe adults, which ensures a pressure gradient for water transport in the more saline environment of the scrub mangrove (Melcher *et al.* 2001, Naidoo 2010, Reef & Lovelock 2015). In contrast, in seedlings, no differences among scrub and fringe were observed, but seedling leaf water potential was higher than that for adults. The higher water potential in seedlings could mean that at this stage, both propagule and leaves are important water reservoirs (Lechthaler *et al.* 2016). The access to water reserves could also explain the similarity in the xylem structure and hydraulic conductivity at both sites for the seedling stage (Ball 1988, Melcher *et al.* 2001, Hao *et al.* 2009). However, the water transport strategies could change for seedlings as they grow, as it has been observed by Kodikara *et al.* (2017) in seedlings of six mangrove species, implying that adaptation to salt and physiological needs of mangrove seedlings vary with age.

Differences in the hydraulic architecture of the two studied stages of scrub *R. mangle* highlight the relevance of the propagule itself and the growing season in which it is established. It has been reported that the balance of reserves and photosynthetic function of the propagule can change depending on the environment in which seedlings develops (Smith & Snedaker 2000). Thus, seedlings that grow in lower salinity conditions, such as those of the fringe forest, can waive the propagule reserves and obtain carbon from photosynthesis (both from the propagule green tissue and from its developing leaves). In contrast, in places where environmental conditions limit photosynthesis and hydraulic conductivity (Naidoo 2006, 2010), such as the scrub mangrove, seedlings can use these reserves for their growth. Furthermore, strategies regarding hydraulic architecture in seedlings could depend on the specific season in which they were established (Dissanayake *et al.* 2014, Schreel *et al.* 2019, Guillén-Rivera *et al.* 2021). These results also highlight the importance of a correct choice or season for reforestation on restoration projects and the importance of seasonal influx of water for successful seedling establishment.

It is possible that at the seedling stage, specific hormonal signals are generated according to the microenvironment and, consequently, these signals can be critical for the formation of a conduction system that guarantees an efficient or safe water flow once the seedling has matured, and the hypocotyl reserves have been exhausted (Farnsworth & Ellison 1996, Farnsworth 2004). Developing an adequate hydraulic architecture according to the microenvironment is essential to survive in fluctuating environmental conditions such as those found in mangrove forests (Ball 1988, 2002, Yan *et al.* 2007, Wang *et al.* 2011). Thus, fringe seedlings ultimately develop an efficient conduction system that allows larger trees to grow larger vessels and greater hydraulic conductivity, while scrub seedlings maintain a safe conduction system throughout their lifetime, even at the cost of limiting their growth.

Although salinity was the most important factor in this study for the differences in hydraulic architecture for plants in the two sites, other factors such as hydroperiod or nutrient availability, which play a significant role in the development of functional strategies of mangrove species (Feller *et al.* 2010) were not considered and should be explored in further studies. A greater understanding of the hydraulic architecture at different life stages allows the development of more exact models on the structure and dynamics of mangrove forests (Ellison & Farnsworth 1997, Vargas-Cruz *et al.* 2019, Peters *et al.* 2020), which can be beneficial for diagnosing their health and for management actions relevant to their restoration and conservation.

## Acknowledgements

We thank J. Luis Simá for help in the field, to Comisión Nacional de Áreas Naturales Protegidas for authorization for sampling in the study site, and to Siuling Castro for providing <u>Figure 1</u>. We also appreciate the help of Ted Killas for English editing. DJCC was the recipient of a fellowship by Consejo Nacional de Ciencia y Tecnología (number 345938). We appreciate the contribution of two anonymous reviewers.

#### Literature cited

- Alongi DM. 2002. Present state and future of the world's mangrove forests. *Environmental Conservation* **29**: 331-349. DOI: <a href="https://doi.org/10.1017/S0376892902000231">https://doi.org/10.1017/S0376892902000231</a>
- Baas P, Werker E. Fahn A. 1983. Some ecological trends in vessel characters. *IAWA Bulletin* 4: 14-147. DOI: <a href="https://doi.org/10.1163/22941932-90000407">https://doi.org/10.1163/22941932-90000407</a>
- Ball MC 1988. Ecophysiology of mangroves. Trees 2: 129-142. DOI: https://doi.org/10.1007/BF00196018
- Ball MC 2002. Interactive effects of salinity and irradiance on growth: implications for mangrove forest structure along salinity gradients. *Trees* **16**: 126-139. DOI: https://doi.org/10.1007/s00468-002-0169-3
- Ball MC, Farquhar GD. 1984. Photosynthetic and stomatal responses of two mangrove species, *Aegiceras corniculatum* and *Avicennia marina*, to long term salinity and humidity conditions. *Plant Physiology* **74**: 1-6. DOI: <a href="https://doi.org/10.1104/pp.74.1.1">https://doi.org/10.1104/pp.74.1.1</a>
- Ball MC, Cochrane MJ, Rawson MH. 1997. Growth and water use of the mangroves *Rhizophora apiculate* and *R. stylosa* in response to salinity and humidity under ambient and elevated concentrations of atmospheric CO<sub>2</sub>. *Plant, Cell & Environment* **20**: 1158-1166. DOI: <a href="https://doi.org/10.1046/j.1365-3040.1997.d01-144.x">https://doi.org/10.1046/j.1365-3040.1997.d01-144.x</a>
- Biber PD. 2006. Measuring the effects of salinity stress in the red mangrove, *Rhizophora mangle* L. *Journal of Agricultural Research* 1: 001-004.
- Cisneros-de la Cruz DJ, Martínez-Castillo J, Herrera-Silveira J, Yáñez-Espinosa L, Ortiz-García, M, Us-Santamaria R, Andrade JL. 2018. Short-distance barriers affect genetic variability of *Rhizophora mangle* L. in the Yucatan Peninsula. *Ecology and Evolution* 8: 11083-11099. DOI: <a href="https://doi.org/10.1002/ece3.4575">https://doi.org/10.1002/ece3.4575</a>
- Cornelissen JHC, Diez PC, Hunt R. 1996. Seedling Growth, Allocation and Leaf Attributes in a Wide Range of Woody Plant Species and Types. *Journal of Ecology* **84**: 755-765. DOI: <a href="https://doi.org/10.2307/2261337">https://doi.org/10.2307/2261337</a>
- Cruiziat P, Cochard H, Améglio T. 2002. Hydraulic architecture of trees: main concepts and results. *Annals of Forest Science* **59**: 723-752. DOI: https://doi.org/10.1051/forest:2002060
- Dissanayake NP, Madarasinghe SK, Jayatissa LP, Koedam N. 2014. Preliminary study on the propagule dependency of *Rhizophora* seedlings. *Journal of the Department of Wildlife Conservation* **2**: 141-151.
- Donovan LA, Ehleringer JR. 1991. Ecophysiological differences among juvenile and reproductive plants of several woody species. *Oecologia* **86**: 594-597. DOI: <a href="https://doi.org/10.1007/BF00318327">https://doi.org/10.1007/BF00318327</a>
- Duke NC, Pinzon MZS. 1992. Aging *Rhizophora* seedlings from leaf scar nodes: A technique for studying recruitment and growth in mangrove forests. *Biotropica* **24**: 173-186. DOI: <a href="https://doi.org/10.2307/2388671">https://doi.org/10.2307/2388671</a>
- Ellison AM, Farnsworth EJ. 1997. Simulated sea level change alters anatomy, physiology, growth, and reproduction of red mangrove (*Rhizophora mangle* L.). *Oecologia* 112: 435-446. DOI: <a href="https://doi.org/10.1007/s004420050330">https://doi.org/10.1007/s004420050330</a>
- Ewers FW, Ewers JM, Jacobsen AL, López-Portillo J, 2007. Vessel redundancy: Modeling safety in numbers. IAWA J. 28, 373-388. DOI: <a href="https://doi.org/10.1163/22941932-90001650">https://doi.org/10.1163/22941932-90001650</a>
- Farnsworth EJ. 2004. Hormones and shifting ecology throughout plant development. *Ecology* **85**: 5-15. DOI: <a href="https://doi.org/10.1890/02-655">https://doi.org/10.1890/02-655</a>
- Farnsworth EJ, Ellison AM. 1996. Sun-shade adaptability of the red mangrove, *Rhizophora mangle* (Rhizophoraceae): Changes through ontogeny at several levels of biological organization. *American Journal of Botany* **83**: 1131-1143. DOI: https://doi.org/10.2307/2446196
- Feller IC. 1996. Effects of nutrient enrichment on leaf anatomy of dwarf *Rhizophora mangle* L. (red mangrove). *Biotropica* **28**: 13-22. DOI: <a href="https://doi.org/10.2307/2388767">https://doi.org/10.2307/2388767</a>

- Feller IC, Sitnik M. 1996. Mangrove ecology: A manual for a field course. Washington, DC: Smithsonian Institute.
- Feller IC, McKee KM, Whigham DF, O'Neill JP. 2003. Nitrogen vs. phosphorus limitation across an ecotonal gradient in a mangrove forest. *Biogeochemistry* **62**: 145-175. DOI: <a href="https://doi.org/10.1023/A:1021166010892">https://doi.org/10.1023/A:1021166010892</a>
- Feller IC, Lovelock CE, Berger U, McKee KL, Joye SB, Ball MC. 2010. Biocomplexity in mangrove ecosystems. *Annual Review of Marine Sciences* **2**: 395-417. DOI: <a href="https://doi.org/10.1146/annurev.marine.010908.163809">https://doi.org/10.1146/annurev.marine.010908.163809</a>
- Gil-Pelegrín E, Aranda I, Peguero-Pina JJ, Vilagrosa A. 2005. El continuo suelo-planta-atmósfera un modelo integrador de la ecofisiología vegetal. *Investigación Agraria Sistemas y Recursos Forestales* 14: 358-370. DOI: http://dx.doi.org/10.5424/srf/2005143-00927
- Guillén-Rivera BE, **López-Portillo** Guzmán JA, **Pérez-**Sánchez E. 2021. Atributos funcionales de plántulas de mangle bajo condiciones ambientales contrastantes. *Kuxulkab* '27: 37-46. DOI: <a href="https://doi.org/10.19136/kuxulkab.a27n57.3690">https://doi.org/10.19136/kuxulkab.a27n57.3690</a>
- Guet J, Fichot R, Lédée C, Laurans F, Cochard H, Delzon S, Bastien C, Brignolas F. 2015. Stem xylem resistance to cavitation is related to xylem structure but not to growth and water-use efficiency at the within-population level in *Populus nigra* L. *Journal of Experimental Botany* **66**: 4643-4652. DOI: <a href="https://doi.org/10.1093/jxb/erv232">https://doi.org/10.1093/jxb/erv232</a>
- Hao GY, Jones TJ, Luton C, Zhang YJ, Manzane E, Scholz FG, Bucci SJ, Cao KF, Goldstein G. 2009. Hydraulic redistribution in dwarf *Rhizophora mangle* trees driven by interstitial soil water salinity gradients: Impacts on hydraulic architecture and gas exchange. *Tree Physiology* **29**: 697-705. DOI: <a href="https://doi.org/10.1093/treephys/tpp005">https://doi.org/10.1093/treephys/tpp005</a>
- Herrera-Silveira JA. 1994. Spatial heterogeneity and seasonal patterns in a tropical coastal lagoon. *Journal of Coastal Research* **10**: 738-746. DOI: <a href="http://www.jstor.org/stable/4298266">http://www.jstor.org/stable/4298266</a>
- Herrera-Silveira JA, Teutli-Hernández C, Zaldívar-Jiménez A, Pérez-Ceballos R, Cortés-Balán, O, Osorio-Moreno I, Ramírez-Ramírez J, Caamal-Sosa J, Andueza-Briceño MT, Torres R y Hernández-Aranda H. 2014. Programa regional para la caracterización y el monitoreo de ecosistemas de manglar del Golfo de México y Caribe Mexicano: Península de Yucatán. Centro de Investigación y de Estudios Avanzados-Mérida. Informe final SNIB-CONABIO, proyecto No. FN009. México DF. México. <a href="http://www.snib.mx/iptconabio/resource?r=SNIB-FN009">http://www.snib.mx/iptconabio/resource?r=SNIB-FN009</a>. (accessed January10, 2013).
- Ishida A, Yazaki K, Hoe AL. 2005. Ontogenetic transition of leaf physiology and anatomy from seedlings to mature trees of a rain forest pioneer tree, *Macaranga gigantea*. *Tree Physiology* **25**: 513-522. DOI: <a href="https://doi.org/10.1093/tree-phys/25.5.513">https://doi.org/10.1093/tree-phys/25.5.513</a>
- Jiang GF, Goodale UM, Liu YY, Hao GY, Cao KF. 2017. Salt management strategy defines the stem and leaf hydraulic characteristics of six mangrove tree species. *Tree Physiology* **37**: 389-401. DOI: <a href="https://doi.org/10.1093/treephys/tpw131">https://doi.org/10.1093/treephys/tpw131</a>
- Jones HG. 2013. *Plants and microclimate: a quantitative approach to environmental plant physiology*. Cambridge: Cambridge University Press. ISBN: 978-051-1845-72-7
- Kodikara KAS, Jayatissa LP, Huxham M, Dahdouh-Guebas F, Koedam N. 2017. The effects of salinity on growth and survival of mangrove seedlings changes with age. *Acta Botanica Brasilica* **32**: 37-46. DOI: <a href="https://doi.org/10.1590/0102-33062017abb0100">https://doi.org/10.1590/0102-33062017abb0100</a>
- Krauss KW, Lovelock CE, McKee KL, López-Hoffman L, Ewe SML, Sousa WP. 2008. Environmental driver in mangrove establishment and early development: A review. *Aquatic Botany* **89**: 105-127. DOI: <a href="https://doi.org/10.1016/j.aquabot.2007.12.014">https://doi.org/10.1016/j.aquabot.2007.12.014</a>
- Lechthaler S, Robert EMR, Tonné N, Prusova A, Gerkema E, Van As H, Koedam N, Windt CW. 2016. Rhizophoraceae mangrove saplings use hypocotyl and leaf water storage capacity to cope with soil water salinity changes. *Frontiers in Plant Science* 7: 1-13. DOI: <a href="https://doi.org/10.3389/fpls.2016.00895">https://doi.org/10.3389/fpls.2016.00895</a>
- Lin G, Sternberg LDSL. 1992. Comparative study of water uptake and photosynthetic gas exchange between scrub and fringe red mangroves, *Rhizophora mangle* L. *Oecologia* **90**: 399-403. DOI: <a href="https://doi.org/10.1007/BF00317697">https://doi.org/10.1007/BF00317697</a>
- López-Portillo J, Ezcurra E. 2002. Los manglares de México: una revisión. *Madera y Bosques* 8: 27-51. DOI: <a href="https://doi.org/10.21829/myb.2002.801290">https://doi.org/10.21829/myb.2002.801290</a>
- López-Portillo J, Ewers FW, Angeles G. 2005. Sap salinity effects on xylem conductivity in two mangrove species. *Plant, Cell & Environment* 28: 1285-1292. DOI: <a href="https://doi.org/10.1111/j.1365-3040.2005.01366.x">https://doi.org/10.1111/j.1365-3040.2005.01366.x</a>
- Lovelock CE, Feller IC, Ball MC, Engelbrecht BMJ, Ewe ML. 2006. Differences in plant function in phosphorus-and nitrogen-limited mangrove ecosystems. *New Phytologist* 172: 514-522. DOI: <a href="https://doi.org/10.1111/j.1469-8137.2006.01851.x">https://doi.org/10.1111/j.1469-8137.2006.01851.x</a>

- Lugo AE, Snedaker SC. 1974. The Ecology of Mangroves. *Annual Review of Ecology and Systematics* **5**: 39-64. DOI: <a href="https://doi.org/10.1146/annurev.es.05.110174.000351">https://doi.org/10.1146/annurev.es.05.110174.000351</a>
- Lugo AE, Medina E, Cuevas E, Cintrón G, Nieves ENL, Novelli YS. 2007. Ecophysiology of a mangrove forest in Jobos Bay, Puerto Rico. *Caribbean Journal of Science* **43**: 200-219.
- Marks OC. 2007. The causes of variation in tree seedling traits: the roles of environmental selection versus chance. *Evolution* **61**: 455-469. DOI: <a href="https://doi.org/10.1111/j.1742-4658.2007.00021.x">https://doi.org/10.1111/j.1742-4658.2007.00021.x</a>
- McCulloh KA, Sperry JS. 2005. Patterns in hydraulic architecture and their implications for transport efficiency. *Tree Physiology* **25**: 257-267. DOI: https://doi.org/10.1093/treephys/25.3.257
- Melcher PJ, Goldstein G, Meinzer FC, Yount DE, Jones TJ, Holbrook NM, Huang CX. 2001. Water relations of coastal and estuarine *Rhizophora mangle*: Xylem pressure potential and dynamics of embolism formation and repair. *Oecologia* 126: 182-192. https://doi.org/10.1007/s004420000519
- Naidoo G. 2006. Factors Contributing to dwarfing in the mangrove *Avicennia marina*. *Annals of Botany* **97**: 1095-1101. DOI: <a href="https://doi.org/10.1093/aob/mc1064">https://doi.org/10.1093/aob/mc1064</a>
- Naidoo G. 2010. Ecophysiological differences between fringe and dwarf *Avicennia marina* man2groves. *Trees* **24**: 667-673. DOI: <a href="https://doi.org/10.1007/s00468-010-0436-7">https://doi.org/10.1007/s00468-010-0436-7</a>
- Peters R, Walther M, Lovelock C, Jiang J, Berger U. 2020. The interplay between vegetation and water in mangroves: new perspectives for mangrove stand modelling and ecological research. *Wetlands Ecology and Management* **28**: 697-712. DOI: <a href="https://doi.org/10.1007/s11273-020-09733-0">https://doi.org/10.1007/s11273-020-09733-0</a>
- Reef R, Lovelock CE. 2015. Regulation of water balance in mangroves. *Annals of Botany* **115**: 385-395. DOI: <a href="https://doi.org/10.1093/aob/mcu174">https://doi.org/10.1093/aob/mcu174</a>
- Robert EMR, Koedam N, Beeckman H, Schmitz N. 2009. A safe hydraulic architecture as wood anatomical explanation for the difference in distribution of the mangroves *Avicennia* and *Rhizophora*. *Functional Ecology* **23**: 649-657. DOI: https://doi.org/10.1111/j.1365-2435.2009.01551.x
- Rodríguez-Zúñiga MT, Troche-Souza C, Vázquez-Lule AD, Márquez-Mendoza JD, Vázquez-Balderas B, Valderrama-Landeros L, Velázquez-Salazar S, Cruz-López MI, Ressl R, Uribe-Martínez A, Cerdeira-Estrada S, Acosta-Velázquez J, Díaz C. 2013. *Manglares de México/Extensión, Distribución y Monitoreo*. México, DF: Comisión Nacional para el Conocimiento y Uso de la biodiversidad. ISBN: 978-607-8328-02-4
- Rosell JA, Olson ME, Anfodillo, T. 2017. Scaling of xylem vessel diameter with plant size: causes, predictions, and outstanding questions. *Current Forestry Report* **3**: 46-59. DOI: <a href="https://doi.org/10.1007/s40725-017-0049-0">https://doi.org/10.1007/s40725-017-0049-0</a>
- Ruzin SE. 1999. Plant microtechnique and microscopy. USA, New York: Oxford University Press. ISBN: 0-19-508956-1
  Schmitz N, Verheyden A, Beeckman H, Kairo JG, Koedam N. 2006a. Influence of a salinity gradient on the vessel characters of the mangrove species *Rhizophora mucronata*. Annals of Botany 98: 1321-1330. DOI: <a href="https://doi.org/10.1093/aob/mcl224">https://doi.org/10.1093/aob/mcl224</a>
- Schmitz N, Verheyden A, De Ridder F, Beeckman H, Koedam N. 2006b. Hydraulic architecture of the mangrove *Rhizophora mucronata* under different salinity and flooding conditions. *In*: Heinrich I, Gärtner H, Monbaron M, Schleser GH, eds. T*RACE* (*Tree Rings in Archaeology, Climatology and Ecology*), vol. 4: *Proceedings of the Dendrosymposium* 2005. Fribourg: Switzerland, pp. 180-187.
- Schreel JDM, Van de Wal BAE, Hervé-Fernandez P, Boeckx P, Steppe K. 2019. Hydraulic redistribution of foliar absorbed water causes turgor-driven growth in mangrove seedlings. *Plant, Cell & Environment* **42**: 2437-2447. DOI: <a href="https://doi.org/10.1111/pce.13556">https://doi.org/10.1111/pce.13556</a>
- Shiau YJ, Lee SC, Chen TH, Tian G, Chiu CY. 2017. Water salinity effects on growth and nitrogen assimilation rate of mangrove (*Kandelia candel*) seedlings. *Aquatic Botany* 137: 50-55. DOI: <a href="https://doi.org/10.1016/j.aquabot.2016.11.008">https://doi.org/10.1016/j.aquabot.2016.11.008</a>
- Smith SM, Snedaker SC. 2000. Hypocotyl Function in Seedling Development of the Red Mangrove, *Rhizophora mangle* L. *Biotropica* **32**: 677-685. DOI: <a href="https://doi.org/10.1646/0006-3606(2000)032[0677:HFISDO]2.0.CO;2">https://doi.org/10.1646/0006-3606(2000)032[0677:HFISDO]2.0.CO;2</a>
- Sobrado MA. 2007. Relationship of water transport to anatomical features in the mangrove *Laguncularia racemosa* grown under contrasting salinities. *New Phytologist* **173**: 584-591. DOI: <a href="https://doi.org/10.1111/j.1469-8137.2006.01927.x">https://doi.org/10.1111/j.1469-8137.2006.01927.x</a>
- Sobrado MA, Ewe SML. 2006. Ecophysiological characteristics of *Avicennia germinans* and *Laguncularia racemosa* coexisting in a scrub mangrove forest at the Indian River Lagoon, Florida. *Trees* **20**: 679-687. DOI: <a href="https://doi.org/10.1007/s00468-006-0083-1">https://doi.org/10.1007/s00468-006-0083-1</a>

- Sperry JS, Donnelly JR, Tyree MT. 1988. A method for measuring hydraulic conductivity and embolism in xylem. *Plant, Cell & Environment* 11: 35-40. DOI: https://doi.org/10.1111/j.1365-3040.1988.tb01774.x
- Tomlinson P. 2016. The Botany of Mangroves. . Cambridge: Cambridge University Press. ISBN: 978-1-107-08067-6
- Tyree M. 2003. Hydraulic limits on tree performance: transpiration, carbon gain and growth of trees. *Trees* **17**: 95-100. DOI: https://doi.org/10.1007/s00468-002-0227-x
- Tyree M, Ewers F. 1991. The hydraulic architecture of trees and other woody plants. *New Phytologist* **119**: 345-360. DOI: <a href="https://doi.org/10.1111/j.1469-8137.1991.tb00035.x">https://doi.org/10.1111/j.1469-8137.1991.tb00035.x</a>
- Tyree M, Zimmermann MH. 2002. Xylem Structure and the Ascent of Sap. Berlin: Springer. ISBN: 3540433546
- Valladares F, Vilagrosa A, Peñuelas J, Ogaya R, Camarero JJ, Corcuera L, Sisó S, Gil-Pelegrín E. 2004. *Ecología del Bosque Mediterráneo en un Mundo Cambiante*. Madrid: Ministerio de Medio Ambiente, EGRAF, SA. ISBN: 84-8014-552-8
- Vargas-Cruz M, Mori GM, Signori-Müller C, Da Silva CC, Oh D, Dassanayake M, Zucchi MI, Silva-Oliveira R, Pereira-Souza A. 2019. Local adaptation of a dominant coastal tree to freshwater availability and solar radiation suggested by genomic and ecophysiological approaches. *Scientific Reports* 9: 19936. DOI: <a href="https://doi.org/10.1038/s41598-019-56469-w">https://doi.org/10.1038/s41598-019-56469-w</a>
- Wang W, Yan Z, You S, Zhang Y, Chen L, Lin, G. 2011. Mangroves: obligate or facultative halophytes? A review. *Trees* 25: 953-963. DOI: <a href="https://doi.org/10.1007/s00468-011-0570-x">https://doi.org/10.1007/s00468-011-0570-x</a>
- Xiao Y, Jie Z, Wang M, Lin G, Wang W. 2009. Leaf and stem anatomical responses to periodical waterlogging in simulated tidal floods in mangrove *Avicennia marina* seedlings. *Aquatic Botany* **91**: 231-237. <a href="https://doi.org/10.1016/j.aquabot.2009.07.001">https://doi.org/10.1016/j.aquabot.2009.07.001</a>
- Yan Z, Wang W, Tang D. 2007. Effect of different time of salt stress on growth and some physiological processes of *Avicennia marina* seedlings. *Marine Biology* **152**: 581-587. DOI: <a href="https://doi.org/10.1007/s00227-007-0710-4">https://doi.org/10.1007/s00227-007-0710-4</a>
- Yáñez-Espinosa L, Flores J. 2011. A review of sea-level rise effect on mangrove forest species: anatomical and morphological modifications. Chapter 15. *In*: Casalegno S. ed. *Global Warming Impacts Case Studies on the Economy, Human Health and on Urban and Natural Environments*. London: Intechopen, pp. 253-276. ISBN: 978-953-307-785-7
- Yáñez-Espinosa L, Terrazas T, López-Mata L. 2001. Effects of flooding on wood and bark anatomy of four species in a mangrove forest community. *Trees* **15**: 91-97. DOI: <a href="https://doi.org/10.1007/s004680000081">https://doi.org/10.1007/s004680000081</a>
- Zaldívar-Jiménez A, Herrera-Silveira J, Alonzo-Parra D. 2004. Estructura y productividad de los manglares en la reserva de biosfera Ría Celestún, Yucatán, México. *Madera y Bosques Número especial*: 25-35. DOI: <a href="https://doi.org/10.21829/myb.2004.1031264">https://doi.org/10.21829/myb.2004.1031264</a>
- Zaldívar-Jiménez MA, Herrera-Silveira JA, Teutli-Hernández C, Comín FA, Andrade JL, Molina CC, Ceballos RP. 2010. Conceptual framework for mangrove restoration in the Yucatán Peninsula. *Ecological Restoration* **28**: 333-342. DOI: <a href="https://doi.org/10.3368/er.28.3.333">https://doi.org/10.3368/er.28.3.333</a>

Associate editor: Silvia Aguilar Rodríguez

**Author contributions**: DJCC, conceptualization, field and laboratory work, data analysis, writing – original draft; LYE, conceptualization, data analysis, wood anatomy supervision, writing – review and editing; CRG, data analysis, writing – review and editing; RUS, field and laboratory work, data analysis; JLA, conceptualization, data analysis, supervision, writing – original draft.